



## STUDY REPORT

### Study Title

ASTM E1052

Standard Test Method to Assess the Activity of Microbicides against Viruses in Suspension

### Product Identity

Pure & Clean Wound Solutions, Wound Cleanser Clinical Strength

Lot: 01020-1

### Test Microorganism

Human Coronavirus, Strain 229-E, ATCC VR-740

### Study Identification Number

NG14797

### Author

Tamisha Smith, B.S.

### Study Completion Date

08APR2020

### Testing Facility

Microchem Laboratory  
1304 W. Industrial Blvd.  
Round Rock, Texas 78681

### Study Sponsor

Pure & Clean, LLC  
Trent Freeman  
1083 W Kathryn Street, Ste 1  
Nixa, MO 65714



## STUDY REPORT SUMMARY

### General Study Information

Study Title: ASTM E1052 Method  
Standard Test Method to Assess the Activity of  
Microbicides against Viruses in Suspension

Study Identification Number: NG14797

### Test System

Test Microorganism: Human Coronavirus, Strain 229-E, ATCC VR-740

Host Cell(s): MRC-5, CCL-171

Test Substance: Pure & Clean Wound Solutions, Wound Cleanser  
Clinical Strength

Lot Number: 01020-1

Test Substance Receipt Date: 04MAR2020

### Test Parameters

Test Substance Dilution: N/A - Ready to Use

Organic Soil Load: No supplementation of organic soil load  
incorporated into inoculum (at level used to  
propagate virus)

Number of Replicates Per Lot: One

Contact Time(s): 15 seconds and 30 seconds

Exposure Temperature: 26.3°C and 46% Relative Humidity (RH)

Neutralization Method(s): Dilution Method using 2% fetal bovine serum (FBS)  
EMEM

### Study Dates

Experimental Start Date/Time: 28MAR2020 / 1323

Experimental Termination Date/Time: 04APR2020 / 0938

Study Completion Date: 08APR2020



## TEST PROCEDURE

### Summary

- Stock virus was thawed and was not supplemented with an organic soil load.
- Test and virus control substances were dispensed in 9-part equivalent volumes into sterile vessels.
- Test and virus control substances were each inoculated with 1-part equivalent volumes of the test virus.
- The test suspensions were held for the contact time(s) of 15 seconds and 30 seconds, as specified by the Study Sponsor, and then neutralized by ten-fold serial dilutions into the appropriate solution.
- The virus control suspension was neutralized in the same manner as the test suspensions.
- Following neutralization, the viral suspensions were quantified to determine the levels of infectious virus using standard cell culture (e.g. TCID<sub>50</sub>) or plaque assay techniques.
- Assay trays/plates were incubated for the period most suitable for the virus-host cell system (e.g. 7 days).
- After the incubation period, the assay was scored for the presence/absence of test virus and cytotoxic effects. The appropriate calculations were performed (e.g. Spearman-Kärber) to determine viral titers and levels of test substance cytotoxicity, where applicable.
- Log<sub>10</sub> and percent reductions were computed for test suspensions relative to the control suspensions and reported to the Study Sponsor.
- Unless otherwise noted, no modifications to the method were made for this study.

### Study Notes

Total Incubation Time: 6 days 19 hours 45 minutes

## SUCCESS CRITERIA

The following measures are met to ensure the acceptability of virucidal efficacy data:

- The virus titer control demonstrate obvious and or typical cytopathic effects on the monolayers unless a detection method other than cytopathic effect is used.
- Neutralization of the test substance with a low titer (e.g. 1000-5000 infective units) of the test virus is demonstrated.
- Quantification of the test and control parameters are conducted at a minimum of four determinations per dilution.

The product performance criteria follows:

- The log and percent reduction of the test virus following exposure to the test substance are calculated however, there is no minimum reduction level to qualify as "passing" or an "efficacious" product.



## CALCULATIONS AND STATISTICAL ANALYSIS

The TCID<sub>50</sub> (Tissue Culture Infectivity Dose) represents the endpoint dilution where 50% of the cell cultures exhibit cytopathic effects due to infection by the test virus. The endpoint dilution at which 50% of the host cell monolayers exhibit cytotoxicity is termed the Tissue Culture Dose (TCD<sub>50</sub>). The TCID<sub>50</sub>, and TCD<sub>50</sub> was determined using the Spearman-Kärber method and calculated as follows:

Negative logarithm of endpoint titer = [- Log of first dilution inoculated] - [((sum of % mortality at each dilution/100) - 0.5) x Logarithm of dilution]

The result of this calculation is expressed as TCID<sub>50</sub>/0.1 ml (or volume of dilution inoculated) for the test, virus control, and neutralization control and TCD<sub>50</sub>/0.1 ml (or volume of dilution inoculated) for the cytotoxicity control.

### Calculation of the Log Reduction

The log reduction in viral titer was calculated as follows:

Plate Recovery Control Log<sub>10</sub> TCID<sub>50</sub> - Virus-Test Substance Log<sub>10</sub> TCID<sub>50</sub>

### Calculation of the Percent Reduction

The percent reduction in viral titer was calculated as follows:

Percent Reduction = 1 - (C/B) x 100, where:

B = Average TCID<sub>50</sub> of virus in control suspensions.

C = Average TCID<sub>50</sub> of virus in virus-test suspensions.

The presence of any test substance cytotoxicity were taken into account when calculating the log and percent reductions in viral titer.

If multiple virus control and test replicates were performed, the average TCID<sub>50</sub> of each parameter was calculated and the average result used to calculate the log reductions in viral titer.



## RESULTS

**Table 1: Virus Control and Test Results**

		Virus Control	Lot: 01020-1 15 seconds	Lot: 01020-1 30 seconds
<b>Cell Control</b>		0 0 0 0	0 0 0 0	0 0 0 0
<b>Dilution</b>	<b>10<sup>-1</sup></b>	+ + + +	N/A	N/A
	<b>10<sup>-2</sup></b>	+ + + +	0 0 0 0	0 0 0 0
	<b>10<sup>-3</sup></b>	+ + + +	0 0 0 0	0 0 0 0
	<b>10<sup>-4</sup></b>	0 + + +	0 0 0 0	0 0 0 0
	<b>10<sup>-5</sup></b>	0 0 0 0	0 0 0 0	N/A
TCID <sub>50</sub> per 0.1 ml		4.25 Log <sub>10</sub>	≤1.50 Log <sub>10</sub>	≤1.50 Log <sub>10</sub>
Log <sub>10</sub> Reduction per 0.1 ml		N/A	≥2.75 Log <sub>10</sub>	≥2.75 Log <sub>10</sub>
Percent Reduction		N/A	≥99.82%	≥99.82%

**Key:** + = Virus recovered; 0 = Virus not recovered and/or no cytotoxicity observed;  
 T = Cytotoxicity observed

**Table 2: Cytotoxicity and Neutralization Control Results**

		Test Sample 01020-1	
		Neutralization	Cytotoxicity
<b>Cell Control</b>		0 0 0 0	0000
<b>Dilution</b>	<b>10<sup>-1</sup></b>	+ + + +	T T T T
	<b>10<sup>-2</sup></b>	+ + + +	0 0 0 0
	<b>10<sup>-3</sup></b>	+ + + +	0 0 0 0
TCD <sub>50</sub> per 0.1 ml		≤0.50 Log <sub>10</sub>	≤0.50 Log <sub>10</sub>

**Key:** + = Virus recovered; 0 = Virus not recovered and/or no cytotoxicity observed;  
 T = Cytotoxicity observed



## STUDY CONCLUSION

The purpose of the study was to determine the virucidal efficacy of Pure & Clean Wound Solutions, Wound Cleanser Clinical Strength (Lot: 01020-1) against Human Coronavirus, Strain 229-E, ATCC VR-740, at contact times of 15 seconds and 30 seconds and at an exposure temperature of 26.3°C and 46% Relative Humidity (RH).

The Virus Control demonstrated an average viral titer of 4.25 Log<sub>10</sub> TCID<sub>50</sub> per 0.1 ml.

Taking the cytotoxicity and neutralization control results into consideration, the evaluated test substance, Pure & Clean Wound Solutions, Wound Cleanser Clinical Strength, demonstrated a  $\geq 2.75$  Log<sub>10</sub> reduction ( $\geq 99.82\%$ ) in viral titer at 15 seconds; a  $\geq 2.75$  Log<sub>10</sub> reduction ( $\geq 99.82\%$ ) at 30 seconds.

Neutralization Control for all test substances demonstrated that the test substance was neutralized at  $\leq 0.50$  Log<sub>10</sub>.

Test substance cytotoxic effects to the host monolayer were observed at  $\leq 0.50$  Log<sub>10</sub> TCD<sub>50</sub> per carrier for Lot 01020-1.

*The test substance will be disposed of 30 days after the completion of this study, unless otherwise requested by the Study Sponsor.*

*The results of this study apply to the tested substances(s) only. Extrapolation of findings to related materials is the responsibility of the Sponsor.*

*Copyright © Microchem Laboratory, 2020. Reproduction and ordinary use of this study report by the entity listed as "Sponsor" is permitted. Other copying and reproduction of all or part of this document by other entities is expressly prohibited, unless prior permission is granted in writing by Microchem Laboratory.*